

**Technical specifications**

**Sensitivity:** 0.05 ppb

**Detection limit**

Tissue and honey.....0.1 ppb  
Shrimp, fish(some interference)..... 0.2 ppb

**Recovery rate**

Shrimp and fish.....90±10%  
Honey ,chicken meat /liver..... 75±15%

**Cross-reaction rate**

AOZ.....100%  
AHD.....<0.1%  
AMOZ.....<0.1%  
SEM.....<0.1%

**Antibiotics ELISA kits available from ADI:**

DE-100010	Clenbuterol ELISA kit, 96 tests (For Urine, Serum, Feed, Meat, Liver)
DE-100020	Ractopamine ELISA kit, (For Liver, Urine, Feed), 96 tests
DE-100030	Salbutamal ELISA kit, For Urine, Tissue, Feed, Animal Tissue, Aquatic, Honey, Intestine,, 96 tests
DE-100040	Chloramphenicol ELISA kit, 96 tests (For Animal Tissue, Aquatic, Honey, Intestine, Urine, Egg, Milk, Serum)
DE-100050	Florfenicol ELISA kit (For Animal Tissue, Aquatic, Honey), 96 tests
DE-100060	Nitrofurantoin (AMOZ) ELISA kit (For Fish, Shrimp, Honey, Chicken/Liver), 96 tests
DE-100070	Nitrofurantoin (AHD) ELISA kit, (For Fish, Shrimp, Honey, Chicken/Liver), 96 tests
DE-100075	Nitrofurantoin (SEM) ELISA kit (Honey, Fish, Shrimp, Chicken/Liver, Fish/Shrimp), 96 tests
DE-100080	Nitrofurantoin (AOZ) ELISA kit (For Fish, Shrimp, Honey, Chicken/Liver), 96 tests
DE-100090	Sulfonamides Residues (SAs) ELISA kit, (For Chicken/Liver, Pork/Liver, Honey/Egg, Serum/Urine, Milk), 96 tests
DE-100100	Sulfamethazine (SM2) ELISA kit, 96 tests (For Chicken/Liver, Pork/Liver, Honey/Egg, Serum/Urine, Milk)
DE-100110	Sulfamethoxydiazine (SMD) ELISA kit, (For Chicken/Liver, Pork/Liver, Honey/Egg, Serum/Urine), 96 tests
DE-100120	Quinolones (QNS) ELISA kit (For Pork/Liver, Chicken/Liver, Shrimp, Fish, Serum, Honey), 96 tests
DE-100130	Enrofloxacin ELISA kit (For Pork/Liver, Chicken/Liver, Shrimp, Fish, Serum, Honey), 96 tests
DE-100140	Ampicillin ELISA kit, (For Pork/Liver, Chicken, Duck, Shrimp, Fish, Honey, Milk), 96 tests
DE-100150	Benzyl Penicillin ELISA kit, (For Pork/Liver, Chicken, Duck, Shrimp, Fish, Honey, Milk), 96 tests
DE-100160	Tylosin ELISA kit (For Meat, Liver, Honey, Egg), 96 tests
DE-100170	Trenbolone ELISA kit (For Animal Tissue, Aquatic, Urine), 96 tests
DE-100180	Diazepam ELISA kit (For Tissue, Urine, Feed), 96 tests
DE-100190	Diethylstilbestrol (DES) ELISA kit (Fish, Shrimp, Liver, Meat, Feed, Urine), 96 tests
DE-100200	Gentamicin ELISA kit (Chicken/Liver), 96 tests
DE-100210	Streptomycin ELISA kit, 96 tests (Chicken/Liver, Honey, Milk)
DE-100230	Olaquinox ELISA kit (Tissue) 96 tests
DE-100240	Sulfaquin-oxaline ELISA kit, (For Pork/Liver, Honey/Egg, Serum/Urine, Milk), 96 tests

See Details at the web site or Contact ADI

## Nitrofurantoin (AOZ) ELISA KIT

**Cat. #DE-100080**

For Qualitative and Quantitative Determination of AOZ in fish, shrimp, honey, and chicken liver.

**India Contact:**

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## Nitrofuran (AOZ) ELISA KIT Cat. #DE-100080

Kit Components, 96 tests	Cat #
Micro-well coated strip plate (12 strips with 8 removable wells each)	DE-100081
6x standard solution (1 ml each): 0.0 ppb, 0.05 ppb, 0.15 ppb, 0.45 ppb, 1.35 ppb, 4.05 ppb	DE-100082
Enzyme conjugate (12 mL)	DE-100083
Antibody working solution (7 mL)	DE-100084
Substrate A solution (7 mL)	DE-SSA
Substrate B solution (7 mL)	DE-SSB
Stop solution (7 mL)	DE-ST
20x concentrated washing buffer (40 mL)	DE-WB
20x concentrated redissolving solution (50 mL)	DE-SS20
2-Nitrobenzaldehyde (C <sub>7</sub> H <sub>5</sub> NO <sub>3</sub> ) (10mL)	DE-100085
Instruction Manual	M-DE-100080

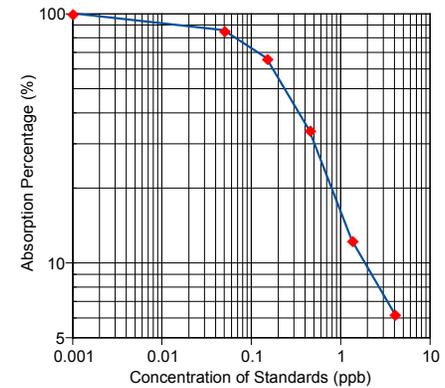
### INTRODUCTION

3-amino-2-oxazolidinone (AOZ) is the metabolite residue of furazolidone. There are four main types of nitrofurans, they are: furazolidone, furaltadone, nitrofurazone and nitrofurantoin.

Furazolidone's chemical formula is C<sub>8</sub>H<sub>7</sub>N<sub>3</sub>O<sub>5</sub> and molecular weight is 225.16 g/mol. It is an antibiotic widely used against Escherichia coli and Salmonella to treat and prevent the infections in gastrointestinal on animals including cattle, swine and poultry. It is also used in veterinary practice as an antibiotic and growth promoter. It can kill various gram-positive and gram-negative bacteria, which are mainly found in animal and aquaculture production. For that reason in some countries it is widely used mixed in animals' water and food. Furazolidone residue, AOZ, can be passed through the food chain to human. AOZ can cause cancer, teratogenesis, and other side effects. The mechanism of the drug works by interfering with various chemical reactions in the bacteria and damaging its DNA. Since its high toxicity on February of 2002 FDA banned the use of any nitrofurans in food-producing animals.

AOZ is very stable in tissue so it will be a good source to be detected on fishes, shrimps and animal muscles and livers. AOZ can be detected by LC-UV, LC-MS, or LC-MS/MS methods, however AOZ Elisa is a more sensitive and low cost tool to detect the use of furazolidone.

Alpha Diagnostic Intl's Nitrofurans (AOZ) ELISA kit is a highly sensitive competitive type assay for the measurement of AOZ in fish, shrimp, honey and chicken liver.



A typical assay Standard Curve (do not use this for calculating sample values)

### CALCULATION OF RESULTS

There are two methods to judge the results: the first one is the rough judgment, while the second is the quantitative determination. Note that the OD value of the sample has a negative correlation with the AOZ concentration.

#### Qualitative determination

The concentration range (ng/mL) can be obtained from comparing the average OD value of the sample with that of the standard solution. Assuming that the OD value of the sample I is 0.268, and that of the sample II is 1.230, the OD value of standard solution is: 1.671 for 0ppb, 1.425 for 0.05ppb, 1.103 for 0.15ppb, 0.567 for 0.45 ppb, 0.205 for 1.35 ppb, 0.104 for 4.05 ppb, accordingly the concentration range of the sample I is 0.45 to 1.35, and that of the sample II is 0.05 to 0.15 ppb.

#### Quantitative determination

The mean values of the absorbance values is obtained for the average OD value (B) of the sample and the standard solution divided by the OD value (B<sub>0</sub>) of the first standard solution (0 standard) and subsequently multiplied by 100%, that is,

$$\text{Percentage of absorbance value} = \frac{B}{B_0} \times 100\%$$

B—the average OD value of the sample or the standard solution  
B<sub>0</sub>—the average OD value of the 0 ng/mL standard solution

Draw the standard curve with the absorption percentages of the standard solution and the semilogarithm values of the AOZ standard solution (ng/mL) as Y- and X-axis, respectively. Read the corresponding concentration of the sample from the standard curve by incorporating its absorption percentage into the standard curve. The resulting value is subsequently multiplied by the corresponding dilution fold, finally obtaining the AOZ concentration in the sample.

Using the professional analyzing software of this kit will be more convenient for the accurate and rapid analysis of a large amount of samples. (Please contact us for this software).

## NOTES:

1. Bring all reagents and micro-well strips to balance at the room temperature (20-25 °C) before use.
2. Return all reagents to 2-8°C immediately after use.
3. The reproducibility of the ELISA analysis, to a large degree, depends on the consistency of plate washing. The correct operation of plate washing is the key point in the procedures of ELISA.
4. For the incubation at constant temperatures, all the samples and reagents must avoid light exposure, and each microplate should be sealed by the cover membrane.
5. The room temperature below 20 °C or the temperature of the reagents and the samples being not returned to the room temperature (20-25 °C) will lead to a lower standard OD value.
6. Dryness of the microplate in the washing process will be accompanied by the situations including the non-linear standard curves and the undesirable reproducibility; So continue to next step immediately after washing.
7. Mix evenly, otherwise there will be the undesirable reproducibility.
8. The stop solution is the 2M sulfuric acid solution, avoid contacting with the skin.
9. Do not use the kit exceeding its expiry date. The use of diluted or adulterated reagents from the kits will lead to the changes in the sensitivity and the detecting OD values. Do not exchange the reagents from the kits of different lot numbers to use.
10. Put the unused microplate into an auto-sealing bag to re-seal it. The standard substance and the colorless color former is light sensitive, and thus they cannot be directly exposed to the light.
11. Discard the coloration solution with any color that indicates the degeneration of this solution. The detecting value of the standard solution 1 ( 0 ppb ) of less than 0.5 indicates its degeneration.

**Work Sheet of Typical Assay-Nitrofurans (AOZ)**

Wells	Stds/samples	Mean A <sub>450 nm</sub>	Absorption Percentage
A1, A2	<b>Standard A</b> 0.0 ppb	1.671	100%
B1, B2	<b>Standard B</b> 0.05 ppb	1.425	85.28%
C1, C2	<b>Standard C</b> 0.15 ppb	1.103	66.01%
D1, D2	<b>Standard D</b> 0.45 ppb	0.567	33.93%
E1, E2	<b>Standard E</b> 1.35 ppb	0.205	12.27%
F1, F2	<b>Standard F</b> 4.05 ppb	0.104	6.22%

NOTE: These data are for demonstration purpose only. A complete standard curve must be run in every assay to determine sample values. Each laboratory should determine their own normal reference values.

## PRINCIPLE OF THE TEST

This test kit is based on the competitive enzyme immunoassay for the detection of AOZ (3-amino-2-oxazolidinone) in the meat, chicken, fish, shrimp and honey. The coupling antigens are pre-coated on the micro-well stripes. The AOZ in the sample and the coupling antigens pre-coated on the micro-well stripes compete for the anti-AOZ antibodies. After the addition of the enzyme conjugate, the TMB substrate is added for coloration. The optical density (OD) value of the sample has a negative correlation with AOZ concentration in the sample. This value is compared to the standard curve and the content of the corresponding AOZ is subsequently obtained.

## MATERIALS AND EQUIPMENT REQUIRED

**Equipments:** microplate reader, printer, vortex, centrifuge, homogenizer, measuring pipettes and balance (a sensibility reciprocal of 0.01 g)

**Micropipettors:** single-channel 20 to 200 µL and 100 to 1000 µL, and multi-channel 250 µL.

**Reagents:** Methanol, NaOH, Ethyl acetate, N-hexane, HCl (approx 36.5%), K<sub>2</sub>HPO<sub>4</sub> 3H<sub>2</sub>O, 2-Nitrobenzaldehyde (C<sub>7</sub>H<sub>5</sub>NO<sub>3</sub>), K<sub>2</sub>Fe(CN)<sub>5</sub>NO 3H<sub>2</sub>O and ZnSO<sub>4</sub> 7H<sub>2</sub>O.

## PRECAUTIONS AND SAFETY INSTRUCTIONS

**The Nitrofurans (AOZ) Kit** is for research use only.

**Stop Solution** contains 1% sulfuric acid. Follow good laboratory practices, and avoid ingestion or contact of any reagent with skin, eyes or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water.

**MSDS for TMB, sulfuric acid**, if not already on file, can be requested or obtained from the ADI website.

## SAMPLE PRE-TREATMENT

### Instructions

The following points must be dealt with before the pre-treatment of any kind of sample:

1. Only the disposable tips can be used for the experiments and the tips must be changed when used for absorbing different reagents.
2. Before the experiment, each experimental equipment must be clean and should be re-cleaned if necessary, in order to avoid the contamination that interferes with the experimental results.

### Solution preparation before sample pre-treatment

1. The 2×concentrated redissolving solution is mixed with deionized water at 1:1 (1 mL 2× concentrated redissolving solution + 1 mL deionized water), used for sample redissolving.

2. C solution (for milk sample): dissolve 12.5 g  $K_2Fe(CN)_5 \cdot NO \cdot 3H_2O$  in deionized water to 100 mL.
3. D solution (for milk sample): dissolve 29.8 g  $ZnSO_4 \cdot 7H_2O$  in deionized water to 100 mL.
4. 0.1 M  $K_2HPO_4$ : dissolve 22.8 g  $K_2HPO_4 \cdot 3H_2O$  in deionized water to 1 L.
5. 1 M HCl: dissolve 8.6 mL HCl (approx 36.5%) in water to 100 mL.
6. 1 M NaOH: dissolve 4 g NaOH in water to 100 mL.

### Samples preparation

#### a) Shrimp, fish and meat

-Homogenize the sample, continue as described in (1 to 7, d)

#### b) Milk

1. Take 5 mL milk into centrifuge tube; add C and D solution, 250  $\mu$ L each
2. Mix thoroughly, use vortex; centrifuge at above 4000 r/min at 4-12  $^{\circ}C$  for 10 min with centrifuge of constant temperatures, if centrifuge of constant temperature is not available, chill sample temperature to approx 8  $^{\circ}C$ , then centrifuge.
3. Continue as described in (1 to 7, d).

#### c) Honey

1. Weigh  $1 \pm 0.05$  g into centrifuge tube.
2. Add 4 mL of the deionized water, mix with vortex; then 0.5 mL 1 M HCl and 100  $\mu$ L 10 mM 2-Nitrobenzaldehyde solution are added, mix thoroughly.
3. Continue as described in (2 to 7, d).

#### d) Continue above steps

1. Weigh  $1 \pm 0.05$  g of the homogenized sample (shrimp, fish or meat), or put 1.1 mL the supernatant of centrifugal milk (equivalent to 1 mL of milk sample) in a plastic tube; add 4 mL of the deionized water, 0.5 mL 1 M HCl and 100  $\mu$ L 10 mM 2-Nitrobenzaldehyde solution to each tube, shake properly.
2. Incubate at 37  $^{\circ}C$  over night ( approx 16 h).
3. Add 5 mL 0.1 M  $K_2HPO_4$ , 0.4 mL 1 M NaOH, 5 mL ethyl acetate to each tube, shake vigorously for 30 s.
4. Centrifuge at above 4000 r/min at room temperature (20-25  $^{\circ}C$ ) for 10 min.
5. Transfer 2.5 mL ethyl acetate layer (upper layer) into a new vessel and evaporate to dryness by nitrogen or air at 50  $^{\circ}C$ .
6. Dissolve the dry residue in 1 mL N-hexane ,add 1 mL of the diluted redissolving solution, and mix properly, centrifuge at above 4000 r/min at room temperature(20-25  $^{\circ}C$ ) for 10 min.
7. Take 50  $\mu$ L of the lower, aqueous phase per well in the assay.

**Fold of dilution of the sample : 2**

#### e) Whole egg

1. Weigh 2 g of the prepared egg sample, put into 50 mL centrifuge tube, add 4 mL of the deionized water, 0.5 mL 1 M HCl, 200  $\mu$ L C solution, mix properly, then add 200  $\mu$ L D solution, shake vigorously for 5 min, centrifuge at above 3000 r/min at room temperature (20-25  $^{\circ}C$ ) for 10 min.

2. Take all clear liquid (upper layer), add 200  $\mu$ L 10 mM 2-Nitrobenzaldehyde solution. Incubate sample in a waterbath for 2 h at 50  $^{\circ}C$  (shake for 1 to 2 min per 0.5 h), add 5 mL 0.1 M  $K_2HPO_4$  and 0.4 mL 1 M NaOH, 5 mL ethyl acetate, shake vigorously for 30 s.
3. Centrifuge at above 4000 r/min at room temperature (20-25  $^{\circ}C$ ) for 10 min.
4. Transfer 2.5 mL of the upper layer into a new vessel and evaporate to dryness by nitrogen or air at 50  $^{\circ}C$ .
5. Dissolve the dry residue in 1 mL n-Hexane, add 2 mL of the diluted redissolving solution, mix properly ,shake for 10 s, centrifuge at above 4000 r/min at room temperature (20-25  $^{\circ}C$ ) for 10 min(if an emulsion forms, place in waterbath at 60  $^{\circ}C$  for up to 5 min until separation occurs), remove the organic phase(upper layer).
6. Take 50  $\mu$ L aqueous phase (the lower) for analysis.

**Fold of dilution of the sample: 2**

### STORAGE AND STABILITY

**Storage:** store at 2 to 8  $^{\circ}C$ , not frozen.

**Expiration date:** 12 months; date of production is on box.

**TEST PROCEDURE (ALLOW ALL REAGENTS TO REACH ROOM TEMPERATURE BEFORE USE).**

### Instructions

1. Bring test kit to the room temperature (20-25  $^{\circ}C$ ) for at least 30 min, note that each reagent must be shaken evenly before use; put the required micro-well strips into plate frames. Resealed the unused microplate, stored at 2-8 $^{\circ}C$ , not frozen.
2. Solution preparation: dilute 40 mL of the concentrated washing buffer (20 $\times$ concentrated) with the distilled or deionized water to 800 mL (or just to the required volume) for use.
3. Numbering: number the micro-wells according to samples and standard solution, each testing sample and standard preparation should be performed in duplicate, record their positions.
4. Add 50  $\mu$ L of the sample or standard solution to separate duplicate wells, and add 50  $\mu$ L of the antibody working solution into each well, seal the microplate with the cover membrane, Mix gently by shaking the plate manually, incubate at 25  $^{\circ}C$  for 1 h.
5. Pour the liquid out of micowell, flap to dry on absorbent paper, add 250  $\mu$ L/well of washing buffer to wash microplate for 10 s, repeat 4-5 times, then take out and flap to dry with absorbent paper.
6. Add 100  $\mu$ L of the enzyme conjugate into each well; and incubate at 25  $^{\circ}C$  for 30 min. Take out microplate, continue as described in 5.
7. Coloration: add 50  $\mu$ L of the substrate A solution and then 50  $\mu$ L of the B solution into each well. Mix gently by shaking the plate manually, and incubate at 25  $^{\circ}C$  for 30 min at dark for coloration.
8. Determination: add 50  $\mu$ L of the stop solution into each well. Mix gently by shaking the plate manually. Set the wavelength of the microplate reader at 450 nm to determine the OD value of every well.(Recommend to read the OD value at the dual-wavelength 450/630 nm within 5 min).